

ASTEROID STEROLS

Masaru Kobayashi, Ryuji Tsuru, Kagemi Todo, and Hiroshi Mitsuhashi

Faculty of Pharmaceutical Sciences, Hokkaido University

Sapporo 060, Japan

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It has been reported by many workers that sterols of asteroids are composed of a mixture of  $\Delta^7$ -mono- and -diunsaturated sterols.<sup>1,2)</sup> The distribution of  $\Delta^7$ -sterols is regarded now as one of the evidences of phylogenetic relationship of asteroids and holothurians.<sup>3)</sup>

Through the investigation of the sterol components of 6 species of asteroids collected in Hokkaido, we found that asteroids generally contain new  $C_{26}$  sterol, asterosterol, as a minor component.<sup>4)</sup> This sterol was also detected in a trace amount in holothurian, Stichopus japonicus by gas liquid chromatography (GLC).

The unsaponifiable matters were obtained by the usual procedure from the asteroids, Asterina pectinifera, Asterias amurensis, Distolasterias sticantha, Certonardoa semiregularis, Lysastrosoma anthosticta, and Solaster paxillatas and the sterol fractions were separated by preparative thin-layer chromatography. Each showed almost the same peaks in GLC (peaks 1 to 7) except for their relative abundances.<sup>5)</sup> Identification of these peaks was made by combined GLC-mass spectrometry (GLC/MS) using free sterols, methyl ether and trimethylsilyl (TMS) ether derivatives from Distolasterias sticantha as a typical one.<sup>6)</sup>

Peak 1 (asterosterol and its methyl and TMS ether) showed molecular ions (M) at  $m/e$  370, 384, 442<sup>7)</sup> and other fragment ions at a (M-Me)  $m/e$  355, 369, 427; b (M-ROH) 352; c (M-Me-ROH) 337; d (M-side chain) 273, 287, -; e (M-side chain-ROH) 255; f (M-side chain-42) 231, 245, -; g (M-side chain-42-ROH) 213; h (M-side chain-27) 246, 260, 318; i (M-side chain-27-RO) 229. The molecular ions and ions a to c show that asterosterol is diunsaturated  $C_{26}$  sterol.

The ions d to i are the common fragments of  $3\beta$ -hydroxy  $\Delta^5$ - and  $\Delta^7$ -sterol and their derivatives<sup>8)</sup>, and accordingly, asterosterol has unsaturated  $C_{7H_{13}}$  side chain. It was confirmed by prominent ions j (M-side chain-2H) at  $m/e$  271, 285 and 343, characteristic of the sterols of unsaturated side chain,<sup>9)</sup> and at  $m/e$  97, derived from side chain fragment.<sup>10)</sup> The ions k (cleavage of C-20 and C-22 with a one hydrogen transfer) at  $m/e$  300, 314 and 372 show that the double bond is located at C-22.<sup>9)</sup> The  $\Delta^5$ -unsaturation was excluded by the absence of ions at  $m/e$  129 and M-129 in TMS ether, and other fragments derived from the fragmentation of A and B rings.<sup>8)</sup> Treatment of 20 $\beta$ -methyl-pregn-7-en- $3\beta$ -ol-21-carboxaldehyde acetate<sup>11)</sup> with isobutyl triphenyl phosphonium bromide in hexane and subsequent hydrolysis afforded 22-cis and -trans mixture of 24-nor-cholesta-5,22-dien- $3\beta$ -ol, resistant to separation. The retention time of more volatile peak agreed with that of asterosterol on 1.5% SE-30 and 1.5% OV-17, showing that asterosterol is 24-nor-cholesta-7,22-dien- $3\beta$ -ol.<sup>12)</sup> Till the present time, we have not succeeded to define the configuration at C-22.

The retention time and cracking pattern of peak 2 agreed with that of the mixture of cholesterol and cholestanol supposed to be of dietary origin.

Other peaks showed sufficient fragments of  $3\beta$ -hydroxy  $\Delta^7$ -sterol ring (ions a to i). The peaks 3 and 5 showed ions j and k compared with ions j and l (M- C-23 to C-27 -1H, characteristic of the  $\Delta^{24}$ -sterols<sup>9)</sup>) at  $m/e$  314, 328, 386 of peaks 6 and 7. From these and their molecular ions, peaks 3 to 7 were identified as shown in the table. It should be pointed out that  $\alpha$ -spinasterol, reported to be obtained by fractional crystallization from Asterina pectinifera and Asterias amurensis,<sup>13)</sup> is not at least a major sterol in these and other species of asteroids studied.

The novel  $C_{26}$  sterol, 22-trans-24-nor-cholesta-5,22-dien- $3\beta$ -ol was isolated first by Idler from mollusc of the class Pelecypoda,<sup>10)</sup> and recently synthesized by Unrau.<sup>14)</sup> We detected the sterol as a minor component in Gastropoda, Littorina brevicula; holothurian, Stichopus japonicus, and annelida of class Polychaeta, Pseudopotamilla ocellata. The sterol was isolated as acetate, mp 141-142<sup>o</sup> (reported,<sup>14)</sup> mp 142.5-143<sup>o</sup>) from Pseudopotamilla ocellata and the spectral data was in good agreement with Idler's  $C_{26}$  sterol. This sterol had a retention time relative to cholestane of 1.20 compared with 1.31 of asterosterol.<sup>5)</sup> The GLC of the sterol mixture of Distolasterias sticantha showed a peak (less than 0.03% of total sterol) whose retention time is identical to this sterol.

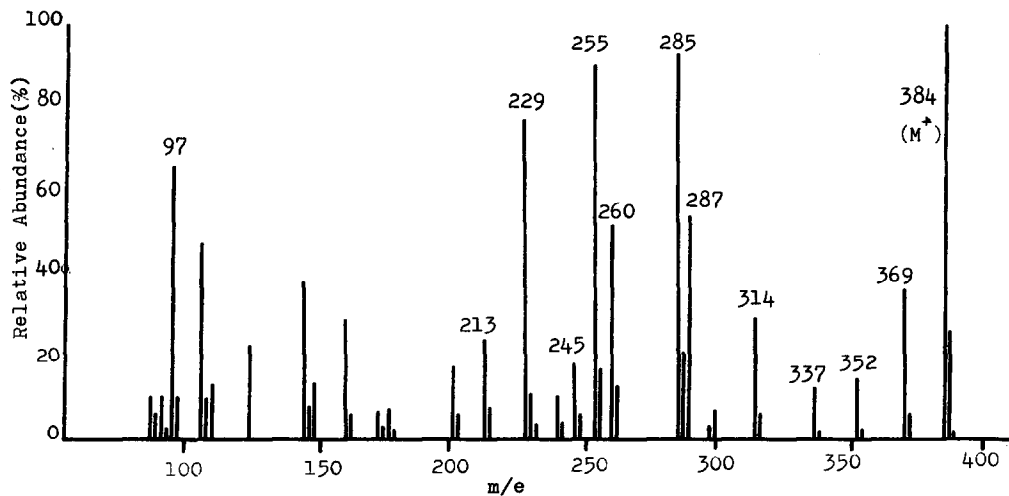
The distribution of  $C_{26}$  sterols in mollusca, arthropoda,<sup>10)</sup> echinodermata, annelida, and protochordata<sup>15)</sup> shows that  $C_{26}$  sterols occur rather widely in marine invertebrates.

TABLE. Components of free sterol

Peak	Relative abundance	Structure	Retention time relative to cholesterol	Mol. wt of sterol and derivatives
1	0.6%	Asterosterol	0.82	370, 384, 442
2	3.5%	Cholesterol Cholestanol	1.00	386, 400, 458 388, 402, 460
3	11.8%	Cholesta-7,22-dien-3 $\beta$ -ol	1.08	384, 398, 456
4	34.2%	Cholest-7-en-3 $\beta$ -ol	1.15	386, 400, 458
5	15.6%	24-Methyl-cholesta-7,22-dien-3 $\beta$ -ol	1.28	398, 412, 470
6	28.2%	24-Methyl-cholest-7-en-3 $\beta$ -ol 24-Methylene-cholest-7-en-3 $\beta$ -ol	1.48	400, 414, 472 398, 412, 470
7	6.1%	24-Ethyl-cholest-7-en-3 $\beta$ -ol 24-Ethylidene-cholest-7-en-3 $\beta$ -ol	1.74	414, 428, 486 412, - , 484

2% OV-17 column, 2 m x 3 mm i.d.; He carrier gas at 30 ml/min; isothermal at 280°.

FIGURE. Mass spectrum of asterosterol methyl ether



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## REFERENCES

1. W. Bergmann, Sterols: Structure and Distribution, in Comparative Biochemistry (M. Florkin and H. S. Mason, Editors) vol. 3; pp.144-152. Academic Press, New York (1962).
2. K.C. Gupta and P.J. Scheuer, Tetrahedron, 24, 5831 (1968).
3. H.I. Bolker, Nature, 213, 904 (1967).
4. This sterol was not detected in GLC of Solaster paxillatas.
5. The comparison of each GLC spectrum was carried out using 1.5% SE-30 column, 1.8 m x 4 mm i.d.; N<sub>2</sub> carrier gas at 60 ml/min; isothermal at 250°.
6. The GLC/MS was carried out using 2% OV-17 column, 2 m x 3 mm i.d.; He carrier gas at 30 ml/min; isothermal at 280° (free), 265° (methyl ether), and 270° (TMS ether).
7. The mass numbers of each sterol derivative are arranged in the order of free sterol, methyl ether, TMS ether in this communication.
8. B.A. Knights, J. Gas Chromatogr., 5, 273 (1967).
9. S.G. Wyllie and C.Djerassi, J. Org. Chem., 33, 305 (1968).
10. D.R. Idler, P.M. Wiseman, and L.M. Safe, Steroids, 16, 451 (1970).
11. K. Sakai and K. Tsuda, Chem. Pharm. Bull. (Tokyo), 11, 529 (1963).
12. While this work was in progress, Goad reported the occurrence of two C<sub>26</sub>-sterols in Henricia sanguinolenta. Although the structure of the side chain of their compounds is uncertain, one of which seems to be identical to asterosterol so far as based on their mass spectral data of TMS ether. L.J. Goad, I. Rubinstein and A.G. Smith, Proc. Roy. Soc. (London), Ser. B., 180, 223 (1972).
13. Y. Toyama and T. Takagi, Bull. Chem. Soc. Japan, 28, 469 (1955).
14. M. Fryberg, A.C. Oehlschlager, and A.M. Unrau, Chem. Commun., 1194 (1971).
15. A. Alcaide, J. Viala, F. Pinte, M. Itoh, T. Nomura, and M. Barbier, Compt. Rend., 273 1386 (1971).